NMR characterisation of five isomeric β,β'-diformyl-mesotetraphenylporphyrins

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Received (in Cambridge, UK) 19th April 2002, Accepted 7th June 2002 First published as an Advance Article on the web 1st July 2002

Five isomeric β , β' -diformyl-meso-tetraphenylporphyrins, and the corresponding nickel complexes, were synthesised, separated and fully characterised by NMR.

Introduction

Formylporphyrins in general, and β-formylporphyrins in particular, are valuable precursors in the synthesis of porphyrin derivatives with important applications in biomedical and natural sciences. 1-3 The introduction of formyl groups into a porphyrin macrocycle is, thus, an important goal for a further functionalization of the macrocycle. The Vilsmeier formylation of nickel complexes of porphyrins is a conventional method for the preparation of these compounds. Porphyrins with meso-unsubstituted positions give meso-formyl derivatives while meso-tetrasubstituted-β-free porphyrins yield β-formyl compounds.⁴⁻⁷ For instance, formylation of (meso-tetraphenylporphyrinato)nickel(II) (1) using POCl₃–DMF in refluxing 1,2dichloroethane gives (β-formyl-meso-tetraphenylporphyrinato)nickel(II) (2a) as the main product 4-6 and a small amount (less than 4%) of a mixture of five nickel complexes of β , β' -diformylmeso-tetraphenylporphyrins (3a-7a). Column chromatography allows an easy separation of the monoformyl derivative from the diformyl ones, which are usually discharged.

During the last years we have been involved in the structural modification of β-formyl-meso-tetraphenylporphyrin derivatives.^{8,9} It was clear for us that each time we were preparing 2a, with the concomitant discharge of the diformyl derivatives, we were wasting potentially useful compounds for further structural modifications. We then decided to set up a project to look for applications of such diformyl compounds as precursors of novel porphyrinic structures. The success of this project depends on how easily the following matters could be solved: a) improvement of the synthesis of the diformyl derivatives, b) the separation of the five isomers and c) their unequivocal structural characterisation. All these questions were successfully approached; we have also shown that these diformyl compounds can be transformed into novel dipyrroloporphyrins.10

In this paper we describe the full details for the synthesis, separation and structural characterisation of the five isomeric β,β'-diformyl-meso-tetraphenylporphyrin derivatives 3a–7a and

Results and discussion

1 Synthesis, separation and nuclear magnetic resonance characterisation of compounds 3a-7a

The diformyl compounds 3a-7a were obtained from 1 by a modification of the standard conditions of the Vilsmeier formylation: a bigger excess of POCl3-DMF and longer reaction times were used. In this way, the diformyl derivatives can be obtained in reasonable amounts. After the hydrolysis of the iminium salts, which are intermediates in the Vilsmeier formylation, and the usual work up, porphyrin 2a (57%) and a mixture of the diformylporphyrins 3a-7a (31%) were obtained. Porphyrin 2a was separated from that mixture by column chromatography; the latter compounds were further separated and purified by preparative thin layer chromatography affording four fractions (these fractions were numbered in order of decreasing R_f). The mass spectra (FAB⁺) of all fractions exhibited the same parent ion at m/z 727. A detailed NMR analysis of reddish fractions 2, 3 and 4 allowed us to identify, respectively, the compounds 3a, 6a and 7a. Fraction 1 (the one with higher R_f and a greenish colour) was shown to be a 1:1 mixture of 4a and 5a (vide infra).

The five diformyl derivatives 3a-7a (Scheme 1) were also obtained, in 71% yield, by direct formylation of the mono-βformyl derivative 2a.

The ¹H NMR spectrum of compound 3a suggests the presence of a symmetric molecule: the resonances of the two formyl protons and the adjacent β-protons appear as two singlets (δ 9.12 and 9.29); the remaining β -protons appear as two doublets (δ 8.66 and 8.74). The HETCOR spectrum of this compound revealed a correlation of the resonance at δ 9.12 with that of the carbon at δ 187.8, and was attributed to the formyl protons. The region corresponding to the ortho protons of the *meso*-phenyl rings revealed three resonances (δ 7.93, 7.96 and 8.00), the latter corresponding to four protons (two equivalent phenyl rings). In the NOESY spectra of 3a the proton resonance of the CHO groups showed a NOE cross peak with the *ortho* protons of the two equivalent phenyl rings (δ 8.00), whereas the singlet corresponding to the β -protons resonance (δ 9.29) showed NOE cross peak with the signals of the protons of another phenyl ring (δ 7.93) (I, Fig. 1). All these data are only compatible with the structure of (2,8-diformyl-mesotetraphenylporphyrinato)nickel(II) 3a. The NOESY spectrum also showed a NOE cross peak between the resonance of the ortho protons of 10,20-phenyl rings and the signal at δ 8.74 and this allowed us to assign it to the resonance of H-12 and H-18

The ¹H NMR spectrum of compound 7a is similar to that of 3a, suggesting the presence of another symmetric diformylporphyrin. The NOESY spectrum of 7a revealed a close proximity between the CHO protons (δ 8.99) and the ortho protons (2 H) of only one phenyl ring (δ 8.07) (III, Fig. 1). These data support the structure of the (2,18-diformyl-mesotetraphenylporphyrinato)nickel(II).

The ¹H NMR spectrum of compound **6a** showed four singlets (δ 9.10, 9.14, 9.24 and 9.32) and four doublets (δ 8.65,

Scheme 1

8.68, 8.71 and 8.73) corresponding to the resonances of the two CHO groups and six β -protons of a porphyrin ring. The HETCOR spectrum of this compound showed correlations between the singlets at δ 9.10 and 9.14 with the carbon resonances at δ 187.7 and 187.8, being assigned to the CHO protons. These NMR data indicate the presence of an asymmetric structure. The NOESY spectra showed close proximities: i) between the *ortho* protons (δ 7.99) of one phenyl ring (5-Ph) and a β -proton (H-3; δ 9.32) and a formyl proton (7-CHO; δ 9.10); ii) between another β -proton (H-8; δ 9.24) and the *ortho* protons (δ 7.93) of another phenyl ring (10-Ph); and also iii) between the other formyl group (2-CHO; δ 9.14) and the *ortho* protons (δ 7.96) of a third phenyl ring (20-Ph) (II, Fig. 1). These data are only compatible with the structure of (2,7-diformyl-meso-tetraphenylporphyrinato)nickel(II) 6a. The analysis of the HMBC spectrum of 6a showed connectivities between H-3 and the carbon resonance at δ 187.8 and also between H-8 and that at δ 187.7, confirming the assignment of those carbon resonances to, respectively, the 2-CHO and 7-CHO groups.

The characterisation of the compound in the first TLC fraction (the one with a greenish colour) was more difficult. Since five isomeric diformyl derivatives were expected and only three of them ($\bf 3a$, $\bf 6a$ and $\bf 7a$) were already found and characterised, the other two expected isomers ($\bf 4a$ and $\bf 5a$) could be anticipated to be present in this fraction. However, only one spot could be observed in the TLC. Its 1H NMR spectrum showed four singlets (δ 9.12, 9.14, 9.28 and 9.29) of identical intensities, the former two being assigned to the resonance of CHO protons (confirmed with the correlations found in their HETCOR spectrum). Our conclusion was that we could have a 1:1 mixture of the two possible isomers $\bf 4a$ and $\bf 5a$. Attempts for their separation by preparative TLC and HPLC were

Fig. 1 Important NOE cross peaks observed in the NOESY spectra of the diformyl derivatives 3a, 6a and 7a.

unsuccessful. Later, we were able to confirm (indirectly) the structures of these two compounds using their free bases **4b** and **5b** (*vide infra*).

2 Synthesis, separation and nuclear magnetic resonance characterisation of compounds 3b-7b

The metal-free forms of the diformyl derivatives 3-7 are, obviously, synthetic targets. Direct acidic demetallation of compounds 3a-7a is not possible due to intramolecular cyclizations which occur during such processes.^{6,11} We were able to promote the demetallation of the intermediate iminium salts resulted from the Vilsmeier formylation of 1 before performing their hydrolysis. The monoformylporphyrin 2b was separated from the diformyl derivatives 3b-7b by column chromatography. A preparative thin layer chromatography of the diformyl derivatives afforded fractions 1-4 (in order of decreasing R_f). Fractions 2, 3 and 4 were identified, respectively, as compounds 3b, 6b and 7b (the same order as in the nickel complexes). Both the ¹H NMR and the HPLC of fraction 1 showed that it is a mixture of two diformylporphyrins in a proportion of ca. 1:1. Careful successive separations of these compounds by preparative TLC afforded small amounts of two fractions consisting of single compounds (confirmed by HPLC). The upper band was identified as 4b and the lower band as **5b**. The ¹H NMR spectrum of the first one (**4b**) indicated the presence of a symmetric structure, since: i) the two formyl groups appear as one singlet (δ 9.18, confirmed by the HETCOR spectrum); ii) the β -protons appear as one singlet (δ 9.38, 2 H) and two doublets (δ 8.93 and 8.96, 2 H each); and iii) the *ortho* protons of the *meso*-phenyl rings also appear as two doublets (δ 8.21 and 8.24, 4 H each). The NOESY spectra of compound **4b** revealed a close proximity between the formyl groups and the *ortho* protons of two phenyl rings (10,20-Ph) appearing at δ 8.24 and also between the β -protons (δ 9.38, H-3 and H-13) with those of the *ortho* protons of the other two phenyl rings (δ 8.21, 5,15-Ph) (**IV**, Fig. 2). These data support the structure of 2,12-diformyl-*meso*-tetraphenylporphyrin **4b**.

Fig. 2 Important NOE cross peaks observed in the NOESY spectra of the diformyl derivatives **4b** and **5b**.

The 1H NMR spectrum of compound **5b** is similar to that of **4b**, however, in this case, the signals corresponding to the CHO and H- β protons resonances appear as four singlets [δ 8.93 (2 H), 8.97 (2 H), 9.17 (2 × CHO) and 9.38 (2 H)], which indicate the presence of the more symmetric diformylporphyrin derivative. The close proximities found in the NOESY spectrum of **5b** (V, Fig. 2) confirm the structure of the expected 2,13-diformyl-*meso*-tetraphenylporphyrin **5b**.

The characterisation of the diformylporphyrins **3b**, **6b** and **7b** was made by comparison of their ¹H NMR spectra with those of the corresponding nickel complexes **3a**, **6a** and **7a**.

Experimental

General

 1 H and 13 C NMR spectra were recorded in deuteriochloroform using Bruker AMX 300 and DRX 300 spectrometers operating at 300.13 and 75.47 MHz, respectively; the spectra of compounds **4b** and **5b** were recorded on a Bruker DRX 500 spectrometer operating at 500.13 and 126.76 MHz, respectively. The chemical shifts are expressed in δ (ppm) values relative to TMS as internal reference and the coupling constants (J) are expressed in Hz. 1 H assignments were made using 2D COSY and NOESY (mixing time of 800 ms) experiments, while 13 C

assignments were made using 2D HETCOR and HMBC experiments (long range C/H coupling constants were optimised to 7 Hz). Mass spectra and HRMS were recorded on VG AutoSpec Q and M mass spectrometers using CHCl₃ as solvent and 3-nitrobenzyl alcohol (NBA) as matrix. Elemental analyses were performed with a Leco CNHS 932 analyzer. The UV–Vis spectra were recorded on a Uvikon spectrophotometer using CH₂Cl₂ as solvent. Column chromatography was carried out using Silica gel (Merck, 35–70 mesh). Preparative thin-layer chromatography was carried out on 20 × 20 cm glass plates coated with Merck 60 silica gel (1 mm thick). Analytical TLC was carried out on precoated sheets with silica gel (Merck 60, 0.2 mm thick).

The HPLC analysis of compounds **4b** and **5b** (retention time 10.2 and 11.0 min, respectively) was carried out in a Gilson Model 305 equipped with a Gilson Model 118 UV–Vis detector and a Spherisorb S5W column using a mixture of ethyl acetate–chloroform–hexane (80: 18: 2) as eluent.

Synthesis of (diformyl-*meso*-tetraphenylporphyrinato)nickel(II) 3a-7a

Method A. A 1,2-dichloroethane (15 ml) solution of 1 (38.6 mg, 0.058 mmol), N,N-dimethylformamide (1.2 ml, 15.8 mmol) and phosphorus oxychloride (0.8 ml, 8.2 mmol) was refluxed for 48 h under a nitrogen atmosphere. The chromatographic analysis (TLC) of the reaction mixture has shown a small amount of the mixture of diformyl compounds (3a-7a). Further portions of N,N-dimethylformamide (1.2 ml) and phosphorus oxychloride (0.8 ml) were added and the resulting mixture was further refluxed for 16 h. The solution was cooled and worked up as described to give 2a (23.0 mg, 57%). The mixture of diformyl compounds was further separated by preparative thin layer chromatography using a (7:3) mixture of dichloromethane-light petroleum as eluent and afforded, in order of increasing polarity, a mixture of greenish compounds 4a and 5a (5.6 mg, 13%), and also the red ones 3a (2.1 mg, 5%), **6a** (4.2 mg, 10%) and **7a** (1.2 mg, 3%).

Method B. A 1,2-dichloroethane (20 ml) solution of **2a** (52.7 mg, 0.076 mmol), *N,N*-dimethylformamide (1.6 ml, 20.7 mmol) and phosphorus oxychloride (1 ml, 10.7 mmol) was refluxed for 24 h under a nitrogen atmosphere. The solution was cooled and worked up to give the starting material **2a** (7.5 mg, 14%), and a mixture of green products **4a** and **5a** (14.1 mg, 26%), and the red ones **3a** (6.9 mg, 13%), **6a** (13.9 mg, 25%) and **7a** (4.0 mg, 7%).

Synthesis of diformyl-meso-tetraphenylporphyrins 3b-7b

N,N-Dimethylformamide (1.2 ml, 15.8 mmol) and phosphorus oxychloride (0.8 ml, 8.2 mmol) were added to a 1,2dichloroethane (15 ml) solution of 1 (38.6 mg, 0.058 mmol). The reaction mixture was refluxed for 48 h under a nitrogen atmosphere. Further portions of N,N-dimethylformamide (1.2 ml) and phosphorus oxychloride (0.8 ml) were added and the resulting mixture was refluxed for another 16 h period. After cooling to room temperature, concentrated H₂SO₄ (6 ml) was added to the reaction mixture, which was stirred for about 10 min at room temperature. The resulting green mixture was washed with water and then with a saturated aqueous solution of sodium carbonate and stirred at room temperature for about 14 h. The organic phase was separated and worked up as described to give 2b (19.3 mg, 52%), a mixture of green compounds 4b and 5b (2.6 mg, 7%) and the red ones 3b (1.2 mg, 3%), **6b** (2.8 mg, 7%) and **7b** (0.6 mg, 2%).

(2,8-Diformyl-*meso*-tetraphenylporphyrinato)nickel(II) 3a. 1 H NMR δ 7.66–7.76 (m, 12 H, H_{meta,para}-Ph), 7.93 (dd, 2 H, J 1.5 and 7.8, H_{ortho}-Ph-5), 7.96 (dd, 2 H, J 2.5 and 6.2, H_{ortho}-Ph-15), 8.00 (dd, 4 H, J 1.4 and 7.9, H_{ortho}-Ph-10,20), 8.66 (d, 2 H, J 5.1,

H-13 and H-17), 8.74 (d, 2 H, *J* 5.1, H-12 and H-18), 9.12 (s, 2 H, 2 × C*H*O), 9.29 (s, 2 H, H-3 and H-7). 13 C NMR δ 118.7, 119.3, 124.7, 125.1; 127.25 and 127.28 (C_{meta}-Ph-5,15), 127.7 (C_{meta}-Ph-10,20); 128.2, 128.6 and 129.2 (C_{para}-Ph); 133.3 (C-13 and C-17), 133.7 (C_{ortho}-Ph-5,15), 133.8 (C_{ortho}-Ph-10,20), 134.1 (C-12 and C-18), 136.9 (C-3 and C-7); 139.0, 139.1, 139.6, 140.4, 141.8, 142.1, 144.3, 145.1; 187.8 (2 × *C*HO); *m/z* (LSIMS) 727 (M + H)⁺.

2,8-Diformyl-meso-tetraphenylporphyrin 3b. ¹H NMR δ –2.15 (s, 2 H, 2 × N*H*), 7.74–7.85 (m, 12 H, H_{meta,para}-Ph), 8.15 (dd, 2 H, *J* 1.5 and 8.0, H_{ortho}-Ph-5), 8.19 (dd, 2 H, *J* 1.9 and 6.0, H_{ortho}-Ph-15), 8.22 (dd, 4 H, *J* 1.7 and 6.5, H_{ortho}-Ph-10,20), 8.77 (d, 2 H, *J* 4.9, H-13 and H-17), 8.83 (d, 2 H, *J* 4.9, H-12 and H-18), 9.24 (s, 2 H, 2 × C*H*O), 9.40 (s, 2 H, H-3 and H-7); ¹³C NMR (75 MHz, CDCl₃) δ 120.2, 120.5, 125.3, 126.97 and 126.99 (C_{meta}-Ph-5, 15), 127.6 (C_{meta}-Ph-10, 20), 128.1, 128.6 and 129.2 (C_{para}-Ph), 132.2 (C-13, 17), 132.9 (C-12, 18), 134.6 (C_{ortho}-Ph-5), 134.7 (C_{ortho}-Ph-15), 135.8 (C_{ortho}-Ph-10, 20), 136.2 (C-3, 7), 140.4 (C-2, 8), 141.0, 141.3, 141.9, 188.8 (2 × CHO); *m/z* (LSIMS) 671 (M + H)⁺, 670 (M⁺⁺); calc. for C₄₆H₃₀N₄O₂·H₂O: C, 80.21; N, 8.13; H, 4.68; found: C, 80.07; N, 8.11; H, 4.54%; λ_{max} (CH₂Cl₂)/nm (log ε): 671 (3.76), 612 (3.79), 575 (4.05), 535 (4.22), 436 (5.47).

(2,12-Diformyl-*meso*-tetraphenylporphyrinato)nickel(II) 4a and (2,13-diformyl-*meso*-tetraphenylporphyrinato)nickel(II) 5a (1:1 mixture). ¹H NMR δ 7.65–7.76 (m, 2 × 12 H, H_{meta + para}-Ph), 7.92–7.96 (m, 8 H, H_{ortho}-Ph), 7.98–8.02 (m, 2 × 8 H, H_{ortho}-Ph), 8.68, 8.70, 8.72 (3 s, 2 × 4 H, H-β), 9.12 (s, 2 H, C*HO*), 9.14 (s, 2 H, C*HO*), 9.28 (s, 2 H, H-β), 9.29 (s, 2 H, H-β). ¹³C NMR δ 118.6, 118.9, 121.8, 122.2; 127.2, 127.68 and 127.71 (C_{meta}-Ph); 128.4 and 129.2 (C_{para}-Ph); 133.5, 133.57, 133.62 and 133.66 (C-β); 133.66, 133.7, 133.83 and 133.9 (C_{ortho}-Ph); 136.6 and 136.9 (C-β); 139.4, 140.0, 140.3, 140.4, 141.1, 141.4, 142.3, 142.5, 143.7, 143.9, 144.8, 144.9; 187.78 and 187.82 (2 × *C* HO); *m/z* (LSIMS) 727 (M + H)⁺.

2,12-Diformyl-*meso***-tetraphenylporphyrin 4b.** ¹H NMR δ –2.53 (s, 2 H, N*H*), 7.76–7.87 (m, 12 H, $H_{meta,para}$ -Ph), 8.21 (d, 4 H, J 6.6, H_{ortho} -Ph-5,15), 8.24 (d, 4 H, J 6.6, H_{ortho} -Ph-10,20), 8.93 (d, 2 H, J 4.8, H-7 and H-17), 8.96 (d, 2 H, J 4.8, H-8 and H-18), 9.18 (s, 2 H, 2 × C*H*O), 9.38 (s, 2 H, H-3 and H-13). ¹³C NMR δ 120.4, 122.9; 127.0 and 127.6 (C_{meta}-Ph); 128.4 and 129.2 (C_{para}-Ph); 134.8 and 135.1 (C_{ortho}-Ph); 139.5 (C-3 and C-13); 141.2, 141.9; 189.0 (2 × CHO) (because of the small amount of sample available these are the only observable ¹³C signals). HRMS (FAB) m/z for C₄₆H₃₁N₄O₂ (M + H)⁺: calculated 671.2447, found 671.2419; λ_{max} (CH₂Cl₂)/nm (log ε): 679 (4.02), 619 (3.64), 575 (3.74), 532 (4.17), 432 (5.42).

2,13-Diformyl-*meso***-tetraphenylporphyrin 5b.** ¹H NMR δ –2.54 (s, 2 H, NH), 7.75–7.87 (m, 12 H, $H_{meta,para}$ -Ph), 8.19 (d, 4 H, J 6.9, H_{ortho} -Ph-5,10), 8.26 (d, 4 H, J 7.0, H_{ortho} -Ph-15,20), 8.93 (s, 2 H, H-7 and H-8), 8.97 (s, 2 H, H-17 and H-18), 9.17 (s, 2 H, 2 × CHO), 9.38 (s, 2 H, H-3 and H-12). ¹³C NMR δ 120.4, 122.9; 127.0 and 127.6 (C_{meta}-Ph); 128.4 and 129.3 (C_{para}-Ph); 130.9 (C-7,8 and C-17,18); 134.7 and 135.3 (C_{ortho}-Ph); 140.2 (C-3 and C-12); 141.0, 142.1; 188.9 (2 × CHO) (because of the small amount of sample available these are the only observable ¹³C signals). HRMS (FAB) m/z for C₄₆H₃₁N₄O₂ (M + H)⁺: calculated 671.2447, found 671.2427; λ_{max} (CH₂Cl₂)/nm (log ε): 679 (4.01), 619 (3.66), 574 (3.71), 532 (4.19), 436 (5.50).

(2,7-Diformyl-meso-tetraphenylporphyrinato)nickel(II) 6a. 1 H NMR δ 7.65–7.79 (m, 12 H, H_{meta,para}-Ph), 7.93 (dd, 2 H, J 2.0 and 7.8, H_{ortho}-Ph-10), 7.96 (dd, 2 H, J 1.4 and 7.4, H_{ortho}-Ph-20), 7.99 (d, 2 H, J 6.8, H_{ortho}-Ph-5), 8.01 (dd, 2 H, J 1.1 and 6.9, H_{ortho}-Ph-15), 8.65 (d, 1 H, J 5.1, H-β), 8.68 (d, 1 H, J 5.2,

H-β), 8.71 (d, 1 H, J 5.2, H-β), 8.73 (d, 1 H, J 5.1, H-β), 9.10 (s, 1 H, 7-CHO), 9.14 (s, 1 H, 2-CHO), 9.24 (s, 1 H, H-8), 9.32 (s, 1 H, H-3). 13 C NMR δ 118.5, 119.7, 121.2, 122.9; 127.3 and 127.7 (C_{meta}-Ph); 128.2, 128.4, 129.2 and 129.4 (C_{para}-Ph); 133.2, 133.6, 133.8 and 134.0 (C-β); 133.7 and 133.9 (C_{ortho}-Ph); 136.2 (C-8), 136.9 (C-3); 139.3, 139.6, 140.05, 140.08, 140.3, 140.4, 140.7, 141.6, 142.2, 142.5, 143.3, 144.6, 144.7, 145.2; 187.7 (7-CHO), 187.8 (2-CHO); m/z (LSIMS) 727 (M + H) $^+$.

2,7-Diformyl-*meso***-tetraphenylporphyrin 6b.** ¹H NMR δ –2.24 (s, 2 H, 2 × N*H*), 7.74–7.88 (m, 12 H, H_{meta,para}-Ph), 8.17 (dd, 2 H, *J* 1.6 and 6.4, H_{ortho}-Ph-10), 8.19 (dd, 2 H, *J* 2.0 and 7.9, H_{ortho}-Ph-20), 8.22 (dd, 2 H, *J* 1.7 and 7.1, H_{ortho}-Ph-5), 8.24 (dd, 2 H, *J* 1.4 and 6.1, H_{ortho}-Ph-10), 8.78 (d, 2 H, *J* 3.8, H-β), 8.82 (d, 1 H, *J* 3.8, H-β), 8.84 (d, 1 H, *J* 3.8, H-β), 9.18 (s, 1 H, 7-CHO), 9.23 (s, 1 H, 2-CHO), 9.37 (s, 1 H, H-8), 9.42 (s, 1 H, H-3); m/z (LSIMS) 671 (M + H)⁺, 670 (M⁺⁺); HRMS (FAB) m/z for C₄₆H₃₁N₄O₂ (M + H)⁺: calculated 671.2447, found 671.2424; λ_{max} (CH₂Cl₂)/nm (log ε): 674 (3.75), 613 (3.85), 577 (4.08), 537 (4.27), 444 (5.50).

(2,18-Diformyl-meso-tetraphenylporphyrinato)nickel(II) 7a.
¹H NMR δ 7.66–7.85 (m, 12 H, H_{meta,para}-Ph), 7.96 (dd, 6 H, J 1.6 and 7.4, H_{ortho}-Ph-5,10,15), 8.07 (dd, 2 H, J 1.4 and 7.6, H_{ortho}-Ph-20), 8.66 (d, 2 H, J 5.1, H-7,13 or H-8,12), 8.74 (d, 2 H, J 5.1, H-8,12 or H-7,13), 8.99 (s, 2 H, 2 × CHO), 9.25 (s, 2 H, H-3 and H-17).
¹³C NMR δ 117.6, 120.0, 121.7; 127.2 and 127.3 (C_{meta}-Ph); 128.2, 128.3 and 128.4 (C_{para}-Ph); 130.4; 133.3 and 133.7 (C-7,13 and C-8,12); 133.6 (C_{ortho}-Ph-15), 133.8 (C_{ortho}-Ph-10,20), 134.7 (C_{ortho}-Ph-15); 136.2 (C-3 and C-17); 139.4, 139.7, 140.5, 141.2, 141.3, 143.0, 143.8, 145.0; 187.5 (2 × CHO); m/z (LSIMS) 727 (M + H)⁺.

2,18-Diformyl-*meso***-tetraphenylporphyrin 7b.** ¹H NMR δ –2.28 (s, 2 H, 2 × N*H*), 7.68–7.92 (m, 12 H, H_{meta,para}-Ph), 8.18 (dd, 6 H, *J* 1.5 and 7.5, H_{ortho}-Ph-5,10,15), 8.28 (d, 2 H, *J* 7.5, H_{ortho}-Ph-20), 8.80 (d, 2 H, *J* 4.8, H-7,13 or H-8,12), 8.82 (d, 2 H, *J* 4.8, H-8,12 or H-7,13), 9.10 (s, 2 H, 2 × CHO), 9.36 (s, 2 H, H-3 and H-17); m/z (LSIMS) 671 (M + H)⁺, 670 (M⁺⁺); calc. for C₄₆H₃₀N₄O₂·½ H₂O: C, 81.28; N, 8.24; H, 4.60; found: C, 81.10; N, 8.67; H, 4.51%; λ _{max}(CH₂Cl₂)/nm (log ε): 670 (3.94), 613 (3.82), 578 (4.11), 537 (4.19), 442 (5.44).

Acknowledgements

Thanks are due to "Fundação para a Ciência e Tecnologia – FCT" for funding the Organic Chemistry Unit and Project Praxis/2/2.1/QUI/145. One of us (A. M. G. Silva) also thanks FCT for a PhD grant.

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